

University of South Bohemia in České Budějovice
Faculty of Science

**Phylogeny of Brimstone butterflies (genus *Gonepteryx*): The
evolution of colour pattern in UV spectrum and geographical
area**

Master thesis

Bc. et Bc. Dana Hanzalová

Supervisor: RNDr. Zdeněk Faltýnek Fric, PhD.

Consultant: Mgr. Jana Marešová

České Budějovice 2018

HANZALOVÁ D. 2018. Phylogeny of Brimstone butterflies (genus *Gonepteryx*): The evolution of colour pattern in UV spectrum and geographical area. Mgr. Thesis, in English. – 24 p., Faculty of Science, University of South Bohemia, České Budějovice, Czech Republic.

Annotation

Phylogeny, phylogeography and evolution of UV reflecting patterns were studied in 12 species of the genus *Gonepteryx*. Sequences of one mitochondrial (COI) and one nuclear gene (Wingless) were used for phylogenetic analyses and reconstruction of the biogeographical events. The results were later compared with the extent of UV reflecting pattern to construct the ancestral situation and evolution of the UV pattern within the genus.

Prohlašuji, že svoji diplomovou práci jsem vypracovala samostatně pouze s použitím pramenů a literatury uvedených v seznamu citované literatury. Prohlašuji, že v souladu s § 47b zákona č. 111/1998 Sb. v platném znění souhlasím se zveřejněním své diplomové práce, a to v úpravě vzniklé vypuštěním vyznačených částí archivovaných Přírodovědeckou fakultou elektronickou cestou ve veřejně přístupné části databáze STAG provozované Jihočeskou univerzitou v Českých Budějovicích na jejích internetových stránkách, a to se zachováním mého autorského práva k odevzdanému textu této kvalifikační práce. Souhlasím dále s tím, aby toutéž elektronickou cestou byly v souladu s uvedeným ustanovením zákona č. 111/1998 Sb. zveřejněny posudky školitele a oponentů práce i záznam o průběhu a výsledku obhajoby kvalifikační práce. Rovněž souhlasím s porovnáním textu mé kvalifikační práce s databází kvalifikačních prací Theses.cz provozovanou Národním registrem vysokoškolských kvalifikačních prací a systémem na odhalování plagiátů.

V Českých Budějovicích, 18.4.2018

.....

Dana Hanzalová

Declaration & Acknowledgement

As the supervisor of Dana Hanzalová, I hereby declare that she played a major role in the process leading to the manuscript presented here as her Master thesis. Dana conducted all the phylogenetic laboratory work, participated in analyses of both phylogenetic and UV data, and wrote the manuscript. The material was collected by me and our collaborators and the photographs of UV pattern were produced by Pavel Pecháček (Charles University in Prague). Dana thus contributed to the overall work by approximately 65%.

RNDr. Zdeněk Faltýnek Fric, PhD.

I would like to thank my supervisor, Zdeněk Faltýnek Fric, for his time, patience and his infinite kindness. I would like to thank Pavel Pecháček for a great contribution to this study. I would also like to thank my colleagues, especially Jana Marešová, Alena Bartoňová and Michal Rindoš for helping me and never abandoning me in the time of need, my friends and family for their support and my fiancé for always standing by my side. Finally I would like to thank every person not mentioned above who contributed to this study, and the Faculty of Science of University of South Bohemia for allowing me to carry on my research. This study would never exist without all of you. Thank you.

Contents

Abstract	1
Introduction	1
Methods.....	2
Sampling	2
DNA extraction, PCR amplification and sequencing.....	3
Phylogenetic and biogeographical analyses	4
Analysis of ultraviolet reflectance	5
Results.....	6
Phylogenetic analyses	6
Biogeographical analysis	8
Ultraviolet patterns.....	9
Discussion	13
Acknowledgements	14
Literature	15
Appendices.....	19

Abstract

The ultraviolet pattern is a common and well known trait amongst the Lepidoptera. That includes the genus *Gonepteryx*, where the males of majority species possess a certain amount of largely variable UV reflecting pattern on the dorsal surfaces of their wings, which evolutionary history we decided to inspect. We used the sequences of one mitochondrial (COI) and one nuclear gene (Wingless) for reconstruction of phylogeny and biogeography of the genus *Gonepteryx*, the connection with the presence of the UV reflecting pattern and its evolution. We inferred that the genus *Gonepteryx* is monophyletic as well as the origin of the UV reflecting pattern in the genus. The ancestor possessed a moderate UV reflecting area on both forewings and hindwings, which later grew larger, approximately the same size, or secondarily disappear in the descendant species. There is a significant correlation between the variability of the UV pattern and the number of species occurring in an area.

Introduction

Sensitivity to ultraviolet light in animals was for the first time observed in 1882 by John Lubbock, the 1st Baron Avebury (Lubbock 1882). Since then, many studies of ultraviolet perception in animals were published in all major animal groups (Tovée 1995): both vertebrates such as reptiles (Fleishman et al. 1993), birds (Burkhardt 1982, 1989; Bennett & Cuthill 1994), mammals (Jacobs et al. 1991), and invertebrates including spiders (Heiling et al. 2003, 2005), beetles (Pope and Hinton 1977) or butterflies (Lutz 1933a; Mazokhin-Porshnyakov 1957; Nekrutenko 1965; Silberglied and Taylor 1978; Silberglied 1979; Eguchi and Meyer-Rochow 1983; Brunton and Majerus 1995; Kemp 2005). Especially the last group, Lepidoptera, has been an object of particular interest for many researchers through the past decades.

One of the best examined families in butterflies regarding ultraviolet reflection is the family Pieridae (Duponchel, 1835), which was an object of several studies (Brunton 1998; Makino et al. 1952; Mazokhin-Porshniakov 1957; Nekrutenko 1964; Obara & Hidaki 1968; Primož 2011), including genus *Gonepteryx* (Leach, 1815). The ultraviolet reflecting pattern appearing on the wings of male butterflies tend to be consistent in the genus *Gonepteryx* (with the exceptions of few species, such as *Gonepteryx rhamni*; Linnaeus, 1758) and therefore can be used as a taxonomic tool, although not the only one (Kudrna 1975). UV reflectance in *Gonepteryx* genus has been studied by Nekrutenko (1964, 1968, 1970),

Brunton et al. (1996), Pecháček et al. (2014) and it was recently explored also in a monograph by Bozano et al. (2016).

The genus *Gonepteryx* consists of 16 middle-sized to large species (Bozano et al. 2016). Majority of them can be found across the Palearctic region with the exception of *Gonepteryx taiwana* (Paravicini, 1913), *Gonepteryx amintha formosana* (Fruhstorfer, 1908) and *Gonepteryx amintha burmensis* (Tytler, 1926) entering Oriental region in southern Himalayas, southern China and Taiwan, respectively (Bozano et al. 2016). The most widespread and possibly the best known species of the genus is *Gonepteryx rhamni* inhabiting a wide area from Northwestern Africa through Mediterranean and boreal Eurasia to the mountains of Central Asia.

Large variability in UV pattern between different species of the genus *Gonepteryx* and also variability inside of the species *G. rhamni* described by Pecháček et al. (2014) led us to a conclusion that it is a proper time to inspect evolutionary history of the UV reflectance pattern in the genus.

The main object of this study was to construct a phylogenetic tree of *Gonepteryx* genus with possibly all the existing species, examine the phylogenetic and biogeographical relationships across the genus and connect them with the presence of the ultraviolet reflecting pattern and its evolution. We hypothesised that the evolution of the UV patterns correlates with the phylogeny of the genus and that there is a wider range of the UV patterns in the areas with two and more species living sympatrically.

Methods

Sampling

Samples were collected directly or obtained from private collections. We gathered and isolated DNA from a total number of 97 samples of 12 species of the genus *Gonepteryx*: 3 samples of *Gonepteryx acuminata* (C. & R. Felder, 1862), 10 samples of *Gonepteryx amintha* (Blanchard, 1871), 9 samples of *Gonepteryx aspasia* (Menetries, 1859), 2 samples of *Gonepteryx cleobule* (Hübner, 1824), 18 samples of *Gonepteryx cleopatra* (Linnaeus, 1767), 2 samples of *Gonepteryx eversi* (Rehnelt, 1974), 6 samples of *Gonepteryx farinosa* (Zeller, 1847), 3 samples of *Gonepteryx maderensis* (Felder, 1862), 3 samples of *Gonepteryx maxima* (Butler, 1885), 9 samples of *Gonepteryx nepalensis* (Doubleday, 1847), 2 samples of *Gonepteryx palmae* (Stamm 1963), 30 samples of *Gonepteryx rhamni* and 3 samples of *Gonepteryx* sp. which are probably *G. rhamni* females but could not be identified properly

due to the absence of UV reflecting pattern (Appendix 1). The sampling covered majority of the area of the genus *Gonepteryx*.

As outgroups, we selected three species of related genera: *Catopsilia florella* (Fabricius, 1775), *Dercas gobrias* (Hewitson, 1864) and *Eurema lisa* (Boisduval & LeConte, 1829).

DNA extraction, PCR amplification and sequencing

Samples used for the analyses were already desiccated. We extracted the DNA from butterfly legs by grinding the dry tissue and used Genomic DNA Mini Kit (Tissue) (Geneaid) for the extraction, following the instructions from the producer. The isolated DNA was kept in microtubes in approximately -20 °C.

The fragments of DNA were amplified using PCR. We run the PCR for one mitochondrial (cytochrome c oxidase subunit I, COI) and one nuclear (Wingless) marker. We used the protocols described by Wahlberg & Wheat (2008). The primers used for the amplification were: HybLCO-HybHCO and HybRon-HybHCO for the first part of COI (COIa) and HybLepWG1-HybLepWG2 for Wingless (Table 1). The reaction was prepared in 12.5 µl of PPP Mastermix (Top-Bio), 8.6 µl of H₂O, 1 µl of reverse primer, 1 µl of forward primer a 1.9 µl of DNA. The following PCR protocol was used: 5 minutes of 95 °C (beginning); 30 seconds of 94 °C, 30 seconds of 50 °C and 90 seconds of 72 °C (35 cycles); 10 minutes of 72 °C (final extension). The presence of products was tested with gel electrophoresis.

The successful samples were sequenced in one-way in Macrogen, South Korea (<http://www.macrogen.com/eng/index/>). We manually aligned the final sequences in Geneious v. 9.0.4. (Kearse et al. 2012). The total length of each sequence was 487 bp for COI and 412 bp for Wingless.

Table 1. Primers used for PCR amplification.

Marker	Part	Type	Primer	Sequence
COI	1	F	HybLCO	5' GGTCAACAAATCATAAAGATATTGG 3'
	1	R	HybHCO	5' TAAACTTCAGGGTGACCAAAAAATCA 3'
	1	F	HybRon	5' GGAGCYCCWGATATAGCTTTCCC 3'
	1	R	HybHCO	5' TAAACTTCAGGGTGACCAAAAAATCA 3'
Wingless	1	F	HybLepWG1	5' GGARTGYAARTGYCAYGGYATGTCTGG 3'
	1	R	HybLepWG2	5' ACTICGCARCACCARTGGAATGTRCA 3'

Phylogenetic and biogeographical analyses

We used 97 samples of the genus *Gonepteryx* from which we successfully obtained the sequences for both molecular markers and 3 samples of related species as outgroups. We used Partition Finder2 (Lanfear et al. 2016) for the selection of the best substitution model. The selected model for the concatenated dataset was GTR+G (Generalised time-reversible model plus gamma distribution).

For the construction of phylogenetic trees we used Maximum Likelihood (RAxML; Stamatakis 2014) and Bayesian analysis (MrBayes 3.2; Ronquist & Huelsenbeck 2003) with sample frequency = 1000, temperature = 0.2, number of chains = 2 and number of runs = 4; 5,000,000 mcmc generations. The rest of the setting was left as default.

Dating of phylogenetic events was conducted in BEAST 1.8.0. (Drummond et al. 2012). For calibration of the tree, we defined 3 nodes, based on the study by Edger et al. (2015). The first one was the separation of *Gonepteryx* genus and outgroups from the rest of Pieridae family (55,6 mil years; standard deviation = 2,5), the second one was the separation of the branch of *Gonepteryx* and *Dercas* from *Eurema* (38,5 mil years; standard deviation = 1,8), and finally the third one was the separation of *Gonepteryx* genus (34,9 mil years; standard deviation = 1,4). The molecular clock was set to uncorrelated log-normal relaxed clock. The coalescent model was set to Constant. The length of chain was 50,000,000 generations.

For the biogeographical analysis, we defined 7 areas: Canary Islands + Madeira (A), Northwestern Africa (B), Boreal Eurasia (C), Mediterranean (D), Central Asian mountains (E), East Asia (F) and Taiwan (G) (Fig. 1). The reconstruction of ancestral area was computed in RASP 4.0 (Yu et al. 2015) using BioGeoBears script (an R package) which compares several alternative biogeographical scenarios and performs inference of biogeographic history on phylogenies, and also model testing and model choice of the many different possible models (dispersal, vicariance, founder-event speciation, DEC, DIVA, BAYAREA, etc.) (Matzke 2013).

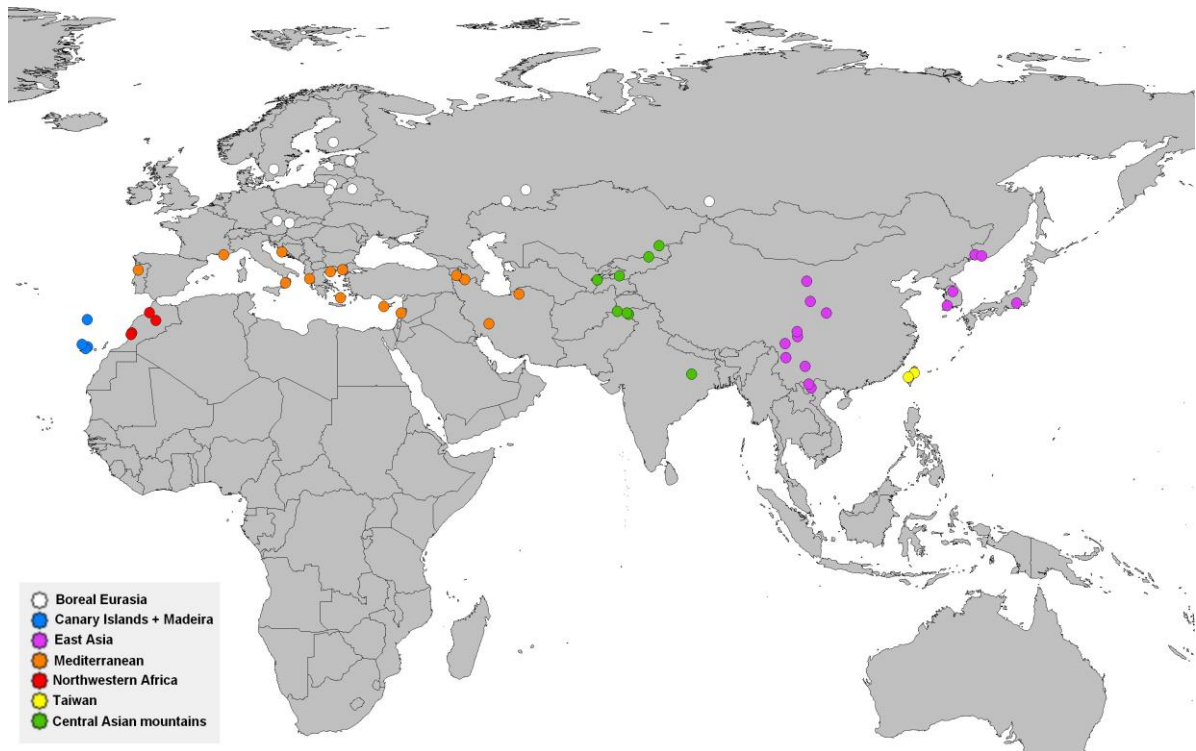


Figure 1. Distribution of *Gonepteryx* spp. samples assigned to the geographical areas used for biogeographical analysis.

Analysis of the ultraviolet reflectance

For the purposes of examining the ancestral situation and evolution of the UV reflectance in *Gonepteryx* genus we put each sample into a table according to presence of the the UV reflectant pattern (present/ absent) and according to the size of the wing area covered with UV reflectant pattern (scale 1-6). The strength of the ultraviolet reflectance was not taken into consideration. The table was prepared separately for the forewings and the hindwings. For taking photographs in the UV wavelength range, we used a FujiFilm IS Pro digital camera suitable for UV photography due to its broad sensitivity spectrum, which spans from 330 to 900 nm. The camera was equipped with an uncoated UV-transmitting lens (Helios 44-2 58mm f/2 lens). We used photographic filters B+W 403 (which blocks the visible spectrum 400–700nm) and B+W BG 53 (which blocks the IR light $\lambda > 700\text{nm}$). As a result, only the UV light ($\lambda < 400\text{nm}$) is transmitted through the lens. For illumination of the photographed objects, we used a UVP MRL-58 multiple-Ray-Lamp (8-watt, 230V-50Hz, 0.16A) equipped with a mercury fluorescent lamp 8w F8T5 long-wave 365nm. All objects were illuminated under the angle of 45° and photographed in a standardized position (dorsal view). The following setting of the FujiFilm IS Pro camera was used for all specimens: ISO

400, shutter time 15', aperture of 3.5. All images were standardized, using 18% gray card, Kodak colour separation guide, and a 15 cm length scale (Pecháček et al. 2014).

Reconstruction of the ancestral state was computed in R 3.3.1 (R Core Team 2013) using fastAnc algorithm (phytools package; Revell 2012), as well as other calculations like simple regressions.

We counted presence of different number of UV patterns per the 7 biogeographical areas and correlated them with a number of species occurring in the area (Table 2). Phylogenetic signal in the UV reflectance was calculated separately for the forewing and the hindwing using Blomberg's K (Blomberg et al. 2007) and Pagel's λ statistics (Pagel 1999). The K values compares variance-covariance effects of the data on the phylogeny with a Brownian motion model. The value of λ is a transformation of the phylogeny that ensures the best fit of trait data to a Brownian Motion mode. In both measures, values close to zero means no phylogenetic signal, whereas values close 1 indicates strong Brownian motion. The values were calculated in R (phylosig in package geiger with 1000 simulations). The values of the variables were compared using Freckleton and Harvey Node-Height Test (nh.test in geiger) (Freckleton & Harvey 2006).

Table 2. Number of species of the genus *Gonepteryx* and number of different types of UV pattern on forewings and hindwings.

Area	Number of species	Different UV patterns on forewing	Different UV patterns on hindwing
Canary Islands + Madeira	2	2	2
Northwestern Africa	2	2	2
Boreal Eurasia	1	1	1
Mediterranean	3	3	3
Central Asian mountains	3	2	1
East Asia	5	4	2
Taiwan	1	1	1

Results

Phylogenetic analyses

The results of the three phylogenetic methods, Maximum Likelihood in RaxML (Appendix 2-4), Bayesian Interference in MrBayes and Beast (Appendix 5) were congruent. The final result is mostly driven by COI since the separate analysis of Wingless is inconclusive and does not reflect the evolution of the studied genus properly. Genus *Gonepteryx* is monophyletic with *Dercas* as the most closely related genus of the chosen outgroups. All species are well defined (Fig. 2).

The origin of *Gonepteryx*, i.e. its split from the common ancestor of *Gonepteryx* and *Dercas*, is estimated around 35 mya (Edger et al. 2015). The first division of *Gonepteryx* genus to the branch containing Eastern Asian species *G. aspasia* and *G. acuminata* and the rest of the species occurred around 22 mya. The rest of Eastern Asian species, namely *G. nepalensis*, *G. amintha* and *G. maxima* separated from the Middle and Western Palearctic branch 16 mya. *G. nepalensis* is a sister species to the other two and the division occurred approximately 10 mya.

The Eurasian group was divided into two main branches 14 mya, when the three species endemic to Canary Islands separated from the branch containing *G. rhamni*, *G. farinosa*, *G. cleopatra* and *G. maderensis*. *G. palmae* is a sister species to *G. eversi* and *G. cleobule* and separated approximately 5 mya.

The most widespread species, *G. rhamni*, separated from the rest of the group around 12 million years ago. Finally, the rest is divided into a branch of *G. farinosa*, which is sister to the branch of *G. cleopatra* and *G. maderensis* and separated 7 mya.

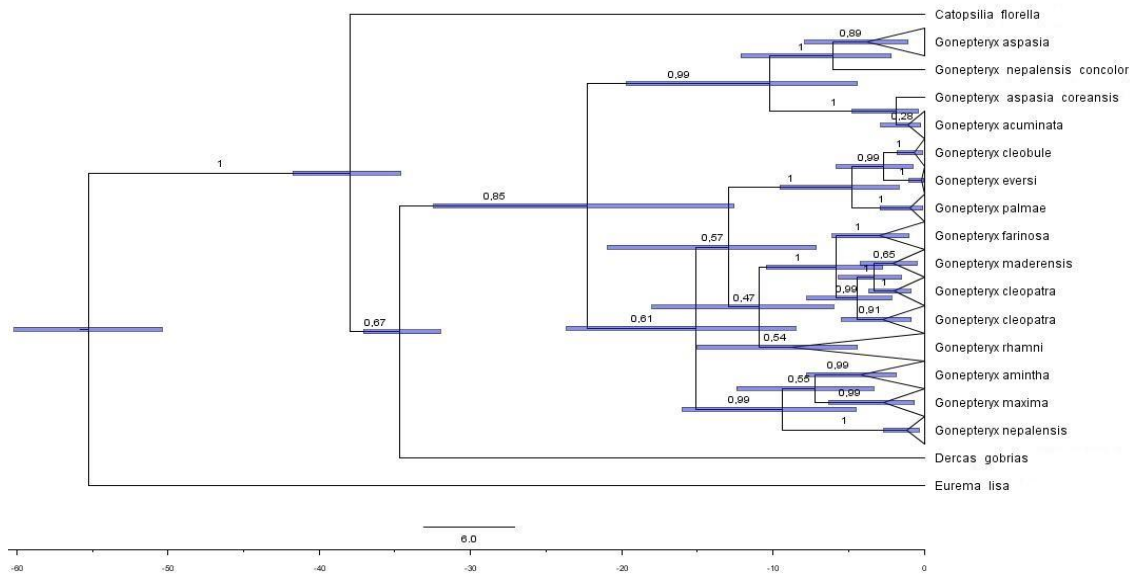


Figure 2. Chronogram showing the phylogeny of the genus *Gonepteryx* based on COI and Wingless and calculated in BEAST 1.8.0. The molecular clock is calibrated by outgroups according to Edger et al. (2015). The branch labels show the posterior probabilities and the time scale is in mya.

Biogeographical analysis

Most favored biogeographical model according the BioGeoBears was DIVALIKE+J (likelihood version of the DIVA model: Ronquist, 1997) (Table 3), which is a likelihood-based model of dispersal-vicariance with additional "j" parameter (founder event/ jump speciation) allowing descendant lineages to have a different area from the direct ancestor (Matzke, 2013; Vasconcelos et al., 2017). Biogeographical analysis (Fig. 3) shows that the common ancestor of *Gonepteryx* genus inhabited a wide area of Central Asian mountains and Eastern Asia including Taiwan and reaching to the Canary Islands. The most frequent biogeographic events were dispersals (26 events) followed by vicariance (18 events). The branch containing *G. aspasia* and *G. acuminata* firstly dispersed into Boreal Eurasia and then separated from the rest of the genus, followed by Central and Eastern Asian branch of *G. amintha* and *G. maxima* and also *G. nepalensis* from Taiwan. The three species show a vicariant occurrence.

The rest of the genus dispersed across the Eurasian area with *G. rhamnii* as the most widespread species with the largest area of occurrence. The group of *G. maderensis*, *G. eversi*, *G. cleobule* and *G. palmae* separated from the rest of the species and remained in the area of the Canary Islands.

Table 3. Parameters of biogeographical models for biogeographical analysis of the genus *Gonepteryx* conducted in BioGeoBears. According to the lowest AIC, the DIVALIKE+J model was selected.

	LnL	numparams	d	e	j	AICc	AICc_wt
DEC	-123.5	2	0.0090	1.0e-12	0	251.1	3.6e-13
DEC+J	-94.7	3	1.0e-12	1.0e-12	0.020	195.6	0.40
DIVALIKE	-117.9	2	0.012	1.0e-12	0	239.8	1.0e-10
DIVALIKE+J	-94.34	3	1.0e-12	1.0e-12	0.020	194.9	0.58
BAYAREALIKE	-168.4	2	0.0086	0.14	0	340.9	1.2e-32
BAYAREALIKE+J	-97.59	3	1.0e-07	1.0e-07	0.021	201.4	0.022

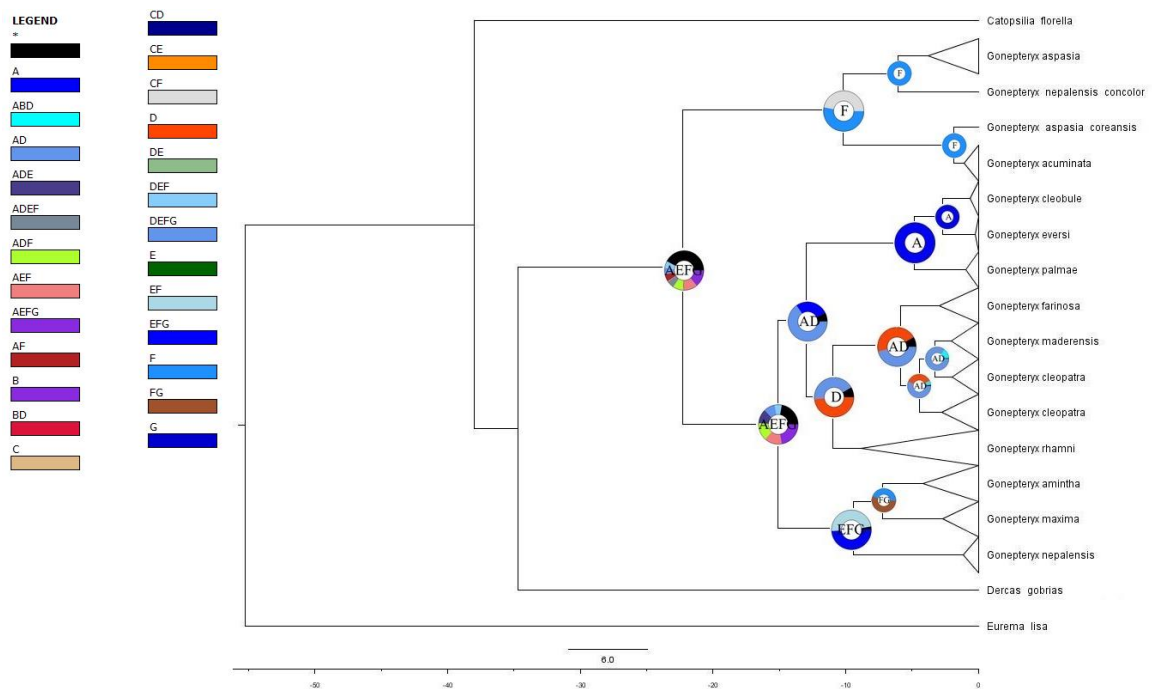


Figure 3. Distribution of the genus *Gonepteryx* in the geographical area in the course of evolution. Time scale is in mya. The regions are (A) Canary Islands + Madeira, (B) Northwestern Africa, (C) Boreal Eurasia, (D) Mediterranean, (E) Central Asian mountains, (F) East Asia and (G) Taiwan.

Ultraviolet patterns

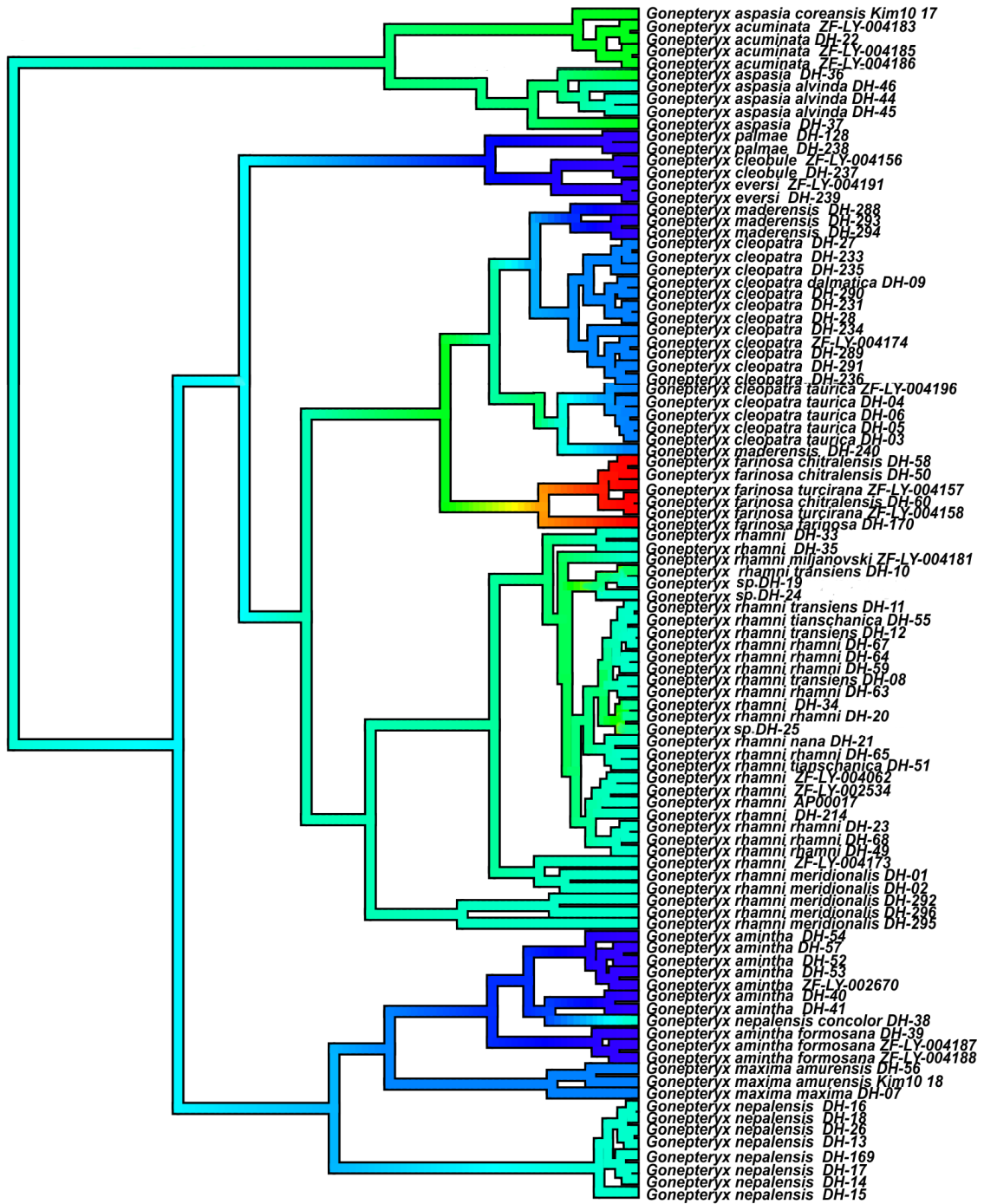
The evolution of ultraviolet patterns in male *Gonepteryx* butterflies shows that the common ancestor possessed a certain amount of ultraviolet reflectance on forewings (Fig. 4) but only a little amount of reflectance on hindwings (Fig. 5).

The surface of forewings covered with ultraviolet pattern grew independently in the branch of Canarian species (*G. maderensis*, *G. eversi*, *G. cleobule* and *G. palmae*) and in Eastern Asian branch of *G. amintha* and *G. maxima*, while in *G. cleopatra* the UV reflecting area grew only a little. The UV reflectance on forewings disappeared completely in *G. farinosa*. For the rest of the species the occurrence and size of UV pattern on forewings did not change.

The presence of UV pattern on hindwings is not as common as on forewings. The most eminent pattern developed in three Canarian species (*G. eversi*, *G. cleobule* and *G. palmae*) while the Madeiran *G. maderensis* and Mediterranean *G. cleopatra* show slightly smaller area of UV reflectance on hindwings but still significantly larger than in the common ancestor. A small area of UV pattern – similar in size to the common ancestor – occurs also in species *G. maxima*, and subspecies *G. amintha formosana* and *G. rhamni meridionalis*. The rest of the species of *Gonepteryx* genus show no sign of UV reflection presence on hindwings.

The relationship between number of species and the variability of the forewing UV pattern was highly significant ($F=53.57$; $df=1, 5$; $p=0.0007$), there was no significant difference in the species number and the variability of the UV pattern on hindwings ($F=1.28$; $df=1, 5$; $p=0.31$).

The Bloomberg's K values for the UV pattern on forewings were equal to 0.261 ($p<0.001$) and for hindwings equal to 2.227 ($p<0.001$). The Pagel's λ values for forewings were 0.95 ($p<<0.001$) and for hindwings 1.004 ($p<<0.001$). It means that the UV pattern has strong phylogenetic signal, however, the low value of K for forewings indicates a presence of a constrain. The UV patterns of both fore- and hindwing were strongly correlated with phylogeny (forewing: intercept=5.30, SE=3.20, $t=1.65$, $p=0.101$, estimate=-0.72, SE=0.16, $t=-4.51$, $p<<0.001$; hindwing: intercept=4.51, SE=2.31, $t=1.96$, $p=0.053$, estimate=-0.73, SE=0.12, $t=-6.38$, $p<<0.001$).



length=11.131

Figure 4. Evolution of UV reflecting pattern in the genus *Gonepteryx* – forewings. Trait value defines the size of UV pattern, where 0 = no UV pattern and 6 = largest area covered with UV pattern.



Figure 5. Evolution of UV reflecting pattern in the genus *Gonepteryx* – hindwings. Trait value defines the size of UV pattern, where 0 = no UV pattern and 6 = largest area covered with UV pattern.

Discussion

We examined the phylogeny of the butterfly genus *Gonepteryx* according to its wing UV reflecting pattern. The study proved that genus *Gonepteryx* is monophyletic and separated from the common ancestor with the species *Dercas* approximately 35 mya (cf. Edger et al. 2015). It is divided into two main groups corresponding with their geographical areas: Eastern and Central Asian group including *G. aspasia*, *G. acuminata*, *G. nepalensis*, *G. amintha* and *G. maxima*; and Eurasian group containing the rest of the species, which could be furthermore divided into the Canarian branch and the rest.

While most species are well defined, there seems to be a problem within a few species. Interesting case is the one of *G. aspasia* and *G. acuminata*. The study of Bozano et al. (2016) works with *G. acuminata* as a subspecies of *G. aspasia*. Our study supports the older interpretation of them being two separate species. However, the situation on the Korean Peninsula has yet to be examined closer since the Korean sample of *G. aspasia* obtained from GenBank clearly belongs to the group of *G. acuminata*. Another problematic species is *G. nepalensis*, which creates two separate branches: *G. nepalensis concolor* and *G. n. nepalensis* in different position. It seems that the subspecies *G. nepalensis concolor* belongs to *G. amintha*. Moreover, our findings that *G. nepalensis* is not a subspecies of *G. rhamnii*, support the findings by Bozano et al. (2016). The situation of *G. cleopatra* is different, according to our data, *G. (cleopatra) taurica* represents a separate species. In addition, the earliest separated branch of *Gonepteryx rhamnii*, i.e. *meridionalis*, may represent a distinct species.

The common ancestor of the genus *Gonepteryx* occurred in a wide area from Eastern Asia including Taiwan and the mountains of Central Asia to Canary Islands. The genus later split up between Asia and the Mediterranean. The genus *Gonepteryx* is most diverse in the terms of species in Asia. Similar pattern can be observed in other butterfly genera, such as *Coenonympha* (Hübner, 1819) studied by Kodandaramaiah & Wahlberg (2009) and also other various groups of flora and fauna (Sanmartín et al., 2001, and references therein). The situation can be explained by the fact that the Pleistocene glaciations were more severe in Europe (which was almost all covered in ice sheets in glacial maxima) than in Asia, causing impoverishment of European biota (Pielou, 1979; Sanmartín et al., 2001). The colonization of Canary Islands and later Mediterranean is rather old and happened before the Messinian Salinity Crisis (7-5.3 mya) when the Mediterranean Sea dried up partially or completely as a result of the closure of its connection with the Atlantic Ocean (Hsü et al., 1973; Krijgsman,

2002; Duggen et al., 2003; Rouchy & Caruso, 2006). However, the refilling of the Mediterranean Sea probably resulted in the vicariant speciation in the Canary Islands which timing correlates with the geographical events.

The origin of UV reflecting pattern in *Gonepteryx* genus is monophyletic. The ancestor possessed a certain amount of UV reflecting pattern on both forewings and hindwings with the UV reflecting area on hindwings being much smaller than on forewings. The UV pattern on both forewings and hindwings is also present in other genera of the family Pieridae, such as *Eurema* Hübner, 1819 (Yata 1989). The UV pattern area later grew significantly larger in the species inhabiting Canary Islands, Mediterranean and Northwestern Africa (both wings) and separately also in the branch containing *G. amintha* and *G. maxima* (forewing only). The UV reflecting pattern on forewings tend to stay the same size or larger during the course of evolution, with the only exception of *G. farinosa* where it disappeared completely. On the other hand, there is rather a disappearing tendency in the hindwing pattern which was already much smaller even in the ancestor.

The study of *G. rhamnii* by Pecháček (2014) declares the possibility of connection between variation in the UV pattern and environmental conditions. However, our study did not confirm this possible pattern on the level of species. The correlation of the forewing UV pattern and the number of species occurring in the area indicates that the UV patterns in the genus *Gonepteryx* might play a role in interspecies communication and recognition.

The possible future studies may concentrate on uncovering relationships between phylogeny and ultraviolet reflectant patterns in other genera of Pieridae family and thus putting this study into a wider context. Another interesting topic might be a deeper exploration of taxonomic position of the few problematic species such as *G. nepalensis*, *G. cleopatra* or the situation of *G. aspasia* and *G. acuminata* on Korean Peninsula.

Acknowledgements

I would like to thank Zdeněk Faltýnek Fric, Pavel Pecháček, Alena Bartoňová, Jana Marešová and Michal Rindoš who significantly contributed to this study. I would also like to thank Layla El Hajj, Vladimír Hula, Josef Jaroš, Jiří Rieger, Vladimír Vrabec, Pavel Vrba & Zdeněk Weidenhoffer, who enabled us to study their collection material. Other material was kindly provided by Alena Bartoňová, Jiří Beneš, Oldřich Čížek, Milan Kopp, Dmitriy V. Morgun, Andro Truuverk, & Michal Zapletal. This research was supported by the Charles University Grant Agency project GAUK 964216.

Literature

BENNETT A. T. D., CUTHILL, I. C. 1994. Ultraviolet vision in birds: What is its function? *Vision Research* 34: 1471–1478.

BLOOMERG S. P., GARLAND T. JR., IVES A. R. 2007. Testing for phylogenetic signal in comparative data: Behavioral traits are more labile. *Evolution* 57: 717–745.

BOZANO G. C., COUTSIS J. G., HEŘMAN P., ALLEGRUCCI G., CESARONI D., SBORDONI V. 2016. Guide to the Butterflies of the Palearctic Region. Pieridae, Part III, Subfamily Coliadinae, Tribes Rhodocerini, Euremini, Coliadini, Genus *Catopsilia* & Subfamily Dismorphiinae. Omnes Artes. Milano. 70 pp.

BRUNTON C. F. A., MAJERUS M. E. N. 1995. Ultraviolet colors in butterflies— intraspecific or inter-specific communication. *Proceedings of the Royal Society B: Biological Sciences* 260: 199–204.

BRUNTON C. F. A., RUSSELL P. J. C., MAJERUS M. E. N. 1996. Variation in ultraviolet wing patterns of brimstone butterflies (*Gonepteryx*: Pieridae) from Madeira and the Canary Islands. *Entomologist* 115: 30–39.

BRUNTON C. F. A. 1998. The evolution of ultraviolet patterns in European *Colias* butterflies (Lepidoptera, Pieridae): A phylogeny using mitochondrial DNA. *Heredity* 80: 611–616.

BURKHARDT D. 1982. Birds, berries and UV. *Naturwissenschaften* 69: 153–157.

BURKHARDT D. 1989. UV vision: a bird's eye view of feathers. *Journal of Comparative Physiology A* 164: 787–796.

DRUMMOND A. J., SUCHARD M. A., XIE D., RAMBAUT A. 2012. Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Molecular Biology and Evolution* 29: 1969–1973.

DUGGEN S., HOERNLE K., BOGAARD P., RUPKE L. 2003. Deep roots of the Messinian salinity crisis. *Nature* 422: 602–606.

EDGER P. P., HEIDEL-FISCHER H. M., BEKAERT M., ROTA J., GLÖCKNER G., PLATTS A. E., HECKEL D. G., DER J. P., WAFULA E. K., TANG M., HOFBERGER J. A., SMITHSON A., HALL J. C., BLANCHETTE M., BUREAU T. E., WRIGHT S. I., dePAMPILIS C. W., ERIC SCHRANZ M., BARKER M. S., CONANT G. C., WAHLBERG N., VOGEL H., PIRES J. C., WHEAT C. W. 2015. The butterfly plant arms-race escalated by gene and genome duplications. *Proceedings of the National Academy of Sciences of the USA* 112: 8362–8366.

EGUCHI E., MEYER-ROCHOW V. B. 1983. Ultraviolet photography of forty three species of Lepidoptera representing ten families. *Annotationes Zoologicae Japonenses* 56: 10–18.

FLEISHMANN L., LEAL M., LOEW E. 1993. Ultraviolet vision in lizards. *Nature* 365: 397–397.

FRECKLETON R. P., HARVEY P. H. 2006. Detecting non-Brownian trait evolution in adaptive radiations. *Public Library of Science Biology* 4: 373.

HEILING A. M., HERBERSTEIN M. E., CHITTKA L. 2003. Pollinator attraction: Crab-spiders manipulate flower signals. *Nature* 421: 334.

HEILING A. M., CHITTKA L., CHENG K., HERBERSTEIN M. E. 2005. Colouration in crab spiders: Substrate choice and prey attraction. *Journal of Experimental Biology* 208: 1785–1792.

HSÜ K. J., RYAN W. B. F., CITA M. B. 1973. Late Miocene desiccation of the Mediterranean. *Nature* 242: 240–244.

JACOBS G. H., NEITZ J., DEEGAN J. F. 1991. Retinal receptors in rodents maximally sensitive to ultraviolet light. *Nature, Lond.* 353: 655–656.

KEARSE M., MOIR R., WILSON A., STONES-HAVAS S., CHEUNG M., STURROCK S., BUXTON S., COOPER A., MARKOWITZ S., DURAN C., THIERER T., ASHTON B., MENTJIES P., DRUMMOND A. J. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28: 1647–1649.

KEMP D. J., RUTOWSKI R. L., MENDOZA M. 2005. Colour pattern evolution in butterflies: A phylogenetic analysis of structural ultraviolet and melanic markings in North American sulphurs. *Evolutionary Ecology Research* 7: 133–141.

KODANDARAMAIAH U., WAHLBERG N. 2009. Phylogeny and biogeography of *Coenonympha* butterflies (Nymphalidae: Satyrinae) – patterns of colonization in the Holarctic. *Systematic entomology* 34: 315–323.

KRIJGSMAN W. 2002. The Mediterranean: Mare Nostrum of Earth sciences. *Earth and Planetary Science Letters* 205: 1–12.

KUDRNA O. 1975. A revision of the genus *Gonepteryx* Leach (Lep., Pieridae). *Entomologist's Gazette* 26: 3–37.

LANFEAR R., CALCOTT B., HO S. Y. W., GUINDON S. 2012. PartitionFinder: Combined selection of partitioning schemes and substitution models for phylogenetic

analyses. *Molecular Biology and Evolution* 29: 1695–1701.

LUBBOCK J. 1882. *Ants, bees, and wasps. A record of observations on the habits of the social Hymenoptera.* D. Appleton and Co., New York

LUTZ F. E. 1933a. Experiments with “stingless bees” (*Trigona cressoni parastigma*) concerning their ability to distinguish ultraviolet patterns. *American Museum Novitates* 641: 1–26.

MAKINO K., SATOH K., UENO N. 1952. Sex in *Pieris rapae* L. and the pteridin content of their wings. *Nature* 170: 933–934.

MATZKE N. 2013. *BioGeoBEARS: BioGeography with Bayesian (and Likelihood) evolutionary analysis in R scripts.* University of California, Berkeley, Berkeley, CA.

MAZOKHIN-PORSHNYAKOV G. A. 1957. Reflecting properties of butterfly wings and the role of ultra-violet rays in the vision of insects. *Biophysics* 2: 285–296.

NEKRUTENKO Y. P. 1964. The hidden wing-patterns of some Palearctic species of *Gonepteryx* and its taxonomic value. *Journal of Research of the Lepidoptera* 3: 65–68.

NEKRUTENKO Y. P. 1965. Gynandromorphic effect and the optical nature of hidden wing-pattern in *Gonepteryx rhamni* L. (Lepidoptera, Pieridae). *Nature* 205: 417–418.

NEKRUTENKO Y. P. 1968. Phylogeny and geographical distribution of the genus *Gonepteryx* (Lepidoptera, Pieridae): An attempt of study in historical zoogeography. *Naukova dumka*, Kiev.

NEKRUTENKO Y. P. 1970. A new subspecies of *Gonepteryx rhamni* from Tian-shan Mountains, U.S.S.R. *Journal of the Lepidopterists' Society* 34: 218–220.

OBARA Y., HIDAKI T. 1968. Recognition of the female by the male, on the basis of ultra-violet reflection, in the white cabbage butterfly, *Pieris rapae crucivora* Boisduval. *Proceedings of the Japan Academy* 44: 829–832.

PAGEL M. 1999. Inferring the historical patterns of biological evolution. *Nature* 401: 877–884.

PECHÁČEK P., STELLA D., KEIL P., KLEISNER K. 2014. Environmental effects on the shape variation of male ultraviolet patterns in the Brimstone butterfly (*Gonepteryx rhamni*, Pieridae, Lepidoptera). *Naturwissenschaften* 101: 1055–1063.

PIELOU E.C. 1979. *Biogeography.* John Wiley & Sons, New York

POPE R. D., HINTON H. E. 1977. A preliminary survey of ultraviolet reflectance in beetles. *Biological Journal of the Linnean Society* 9: 331–348.

PRIMOŽ P., WILTS B. D., STAVENGA D. G. 2011. Spatial reflection patterns of

iridescent wings of male pierid butterflies: curved scales reflect at a wider angle than flat scales. *Journal of Comparative Physiology A* 197: 987–997.

R CORE TEAM. 2013. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

REVELL L. J. 2012. Phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution* 3: 217–223.

RONQUIST F. 1997. Dispersal-vicariance analysis: A new approach to the quantification of historical biogeography. *Systematic Biology* 46: 195–203.

RONQUIST F., HUELSENBECK J. P. 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574.

ROUCHY J. M., CARUSO A. 2006. The Messinian salinity crisis in the Mediterranean basin: A reassessment of the data and an integrated scenario. *Sedimentary Geology* 188–189: 35–67.

SANMARTÍN I., ENGHOFF H., RONQUIST F. 2001. Patterns of animal dispersal, vicariance and diversification in the Holarctic. *Biological Journal of the Linnean Society* 73: 345–390.

SILBERGLIED R. E., TAYLOR O. R. 1978. Ultraviolet reflection and its behavioral role in courtship of sulfur butterflies *Colias eurytheme* and *Colias philodice* (Lepidoptera, Pieridae). *Behavioral Ecology and Sociobiology* 3: 203–243.

SILBERGLIED R. E. 1979. Communication in the ultraviolet. *Annual Review of Ecology, Evolution and Systematics* 10: 373–398.

STAMATAKIS A. 2014. RAxML Version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics*. Open access.

TOVÉE M. J. 1995. Ultra-violet photoreceptors in the animal kingdom: Their distribution and function. *Trends in Ecology & Evolution* 10: 455–459.

WAHLBERG N., WHEAT C. W. 2008. Genomic outposts serve the phylogenomic pioneers: designing novel nuclear markers for genomic DNA extractions of Lepidoptera. *Systematic Biology* 57: 231–242.

YATA O. 1989. A revision of the Old World species of the genus *Eurema* Hübner (Lepidoptera, Pieridae). *Bulletin of the Kitakyushu Museum of Natural History* 9: 1–103.

YU Y., HARRIS A. J., BLAIR C., He X. 2015. RASP (Reconstruct Ancestral State in Phylogenies): A tool for historical biogeography. *Molecular Phylogenetics and Evolution*. 87: 46–49.

Appendices

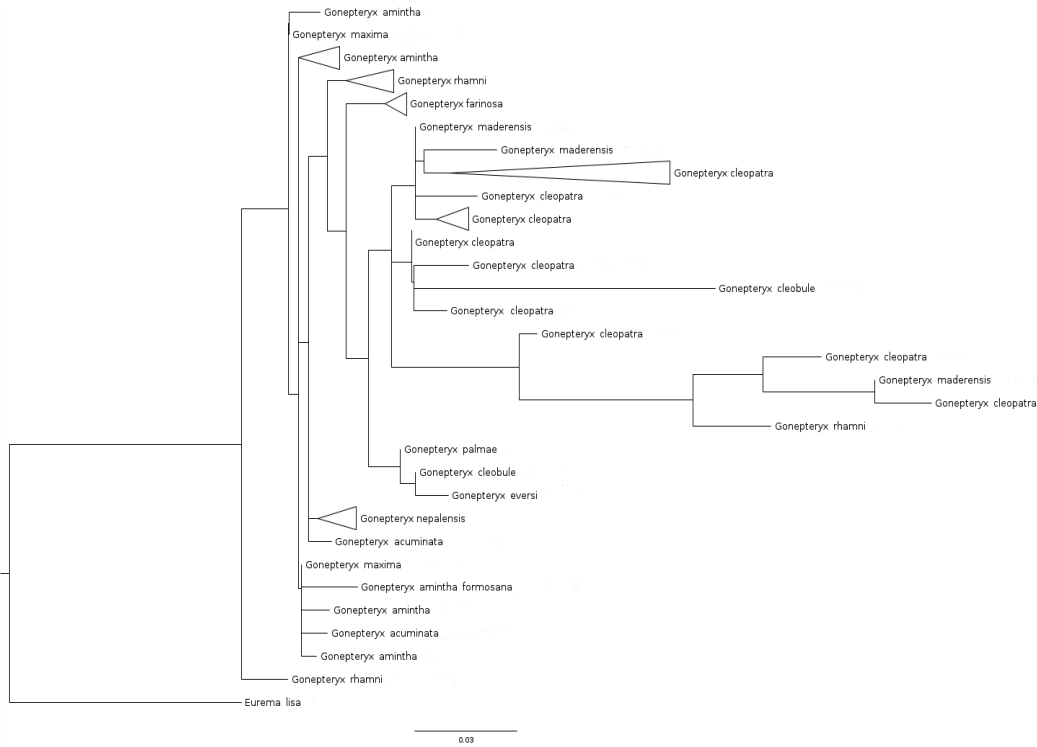
Appendix 1: Table of samples used in the study of wing UV pattern in the genus *Gonepteryx*.

Name	Code	Country	Collection date
<i>G. acuminata</i>	DH-22	Russia	-
<i>G. acuminata</i>	ZF-LY-004185	China	10. 7. 2010
<i>G. acuminata</i>	ZF-LY-004186	China	10. 7. 2010
<i>G. amintha</i>	DH-39	China	28. 4. 2015
<i>G. amintha</i>	DH-40	China	26. 6. 2013
<i>G. amintha</i>	DH-41	China	26. 6. 2013
<i>G. amintha</i>	DH-52	Vietnam	-
<i>G. amintha</i>	DH-53	Vietnam	-
<i>G. amintha</i>	DH-54	Vietnam	-
<i>G. amintha</i>	DH-57	Vietnam	-
<i>G. amintha</i>	ZF-LY-002670	Vietnam	1. 8. 2013
<i>G. amintha</i>	ZF-LY-004187	Taiwan	22. 6. 2016
<i>G. amintha</i>	ZF-LY-004188	Taiwan	1. 5. 2002
<i>G. aspasia</i>	DH-36	China	3. 7. 2009
<i>G. aspasia</i>	DH-44	China	-
<i>G. aspasia</i>	DH-45	China	18. 7. 2007
<i>G. aspasia</i>	DH-46	China	23. 7. 2007
<i>G. aspasia</i>	Kim10_17	Korea	-
<i>G. aspasia</i>	ZF-LY-004183	China	9. 7. 2010
<i>G. cleobule</i>	ZF-LY-004156	Spain	7. 3. 2013
<i>G. cleobule</i>	DH-237	Spain	22. 2. 2010
<i>G. cleopatra</i>	DH-03	Lebanon	27. 5. 2015
<i>G. cleopatra</i>	DH-04	Lebanon	24. 4. 2016
<i>G. cleopatra</i>	DH-05	Lebanon	13. 3. 2016
<i>G. cleopatra</i>	DH-06	Lebanon	9. 5. 2016
<i>G. cleopatra</i>	DH-09	Greece	13. 6. 2008
<i>G. cleopatra</i>	DH-27	Greece	14. 6. 2008
<i>G. cleopatra</i>	DH-28	Greece	13. 6. 2008
<i>G. cleopatra</i>	ZF-LY-004196	Cyprus	26. 5. 2014
<i>G. cleopatra</i>	ZF-LY-004174	Morocco	15. 4. 2010
<i>G. cleopatra</i>	DH-231	Morocco	-

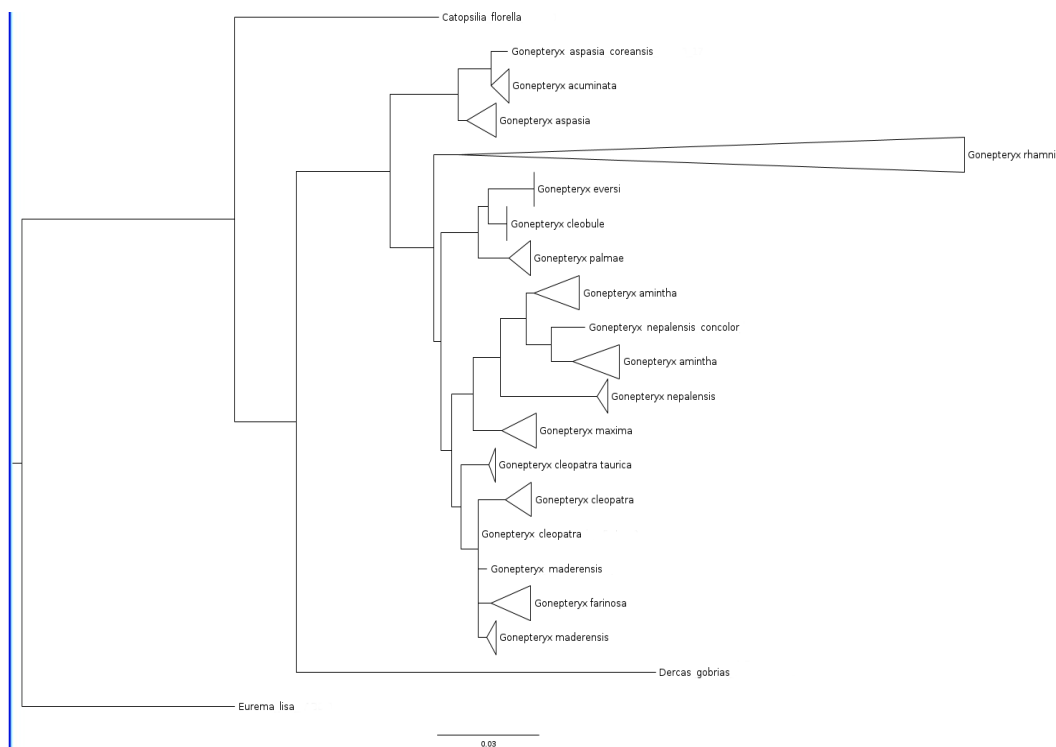
<i>G. cleopatra</i>	DH-233	France	-
<i>G. cleopatra</i>	DH-234	Italy	-
<i>G. cleopatra</i>	DH-235	Croatia	-
<i>G. cleopatra</i>	DH-236	Greece	6. 6. 2012
<i>G. cleopatra</i>	DH-240	Portugal	28. 7. 2011
<i>G. cleopatra</i>	DH-289	Morocco	-
<i>G. cleopatra</i>	DH-290	Morocco	-
<i>G. cleopatra</i>	DH-291	Morocco	-
<i>G. eversi</i>	ZF-LY-004191	Spain	6. 10. 2007
<i>G. eversi</i>	DH-239	Spain	14. 7. 2015
<i>G. farinosa</i>	DH-50	Tajikistan	7. 7. 2015
<i>G. farinosa</i>	DH-58	Kyrgyzstan	5. 7. 2012
<i>G. farinosa</i>	DH-60	Kyrgyzstan	5. 7. 2012
<i>G. farinosa</i>	ZF-LY-004157	Iran	7. 7. 2016
<i>G. farinosa</i>	ZF-LY-004158	Iran	7. 7. 2016
<i>G. farinosa</i>	DH-170	Greece	13. 4. 2011
<i>G. maderensis</i>	DH-288	Madeira	-
<i>G. maderensis</i>	DH-293	Madeira	-
<i>G. maderensis</i>	DH-294	Madeira	-
<i>G. maxima</i>	DH-07	Japan	3. 8. 1997
<i>G. maxima</i>	DH-56	Russia	1. 5. 2015
<i>G. maxima</i>	Kim10_18	Korea	-
<i>G. nepalensis</i>	DH-13	Pakistan	25. 8. 2011
<i>G. nepalensis</i>	DH-14	Pakistan	22. 8. 2011
<i>G. nepalensis</i>	DH-15	Pakistan	26. 8. 2011
<i>G. nepalensis</i>	DH-16	Pakistan	21. 8. 2011
<i>G. nepalensis</i>	DH-17	Pakistan	1. 8. 2011
<i>G. nepalensis</i>	DH-18	Pakistan	28. 8. 2011
<i>G. nepalensis</i>	DH-26	Pakistan	22. 8. 2011
<i>G. nepalensis</i>	DH-38	China	28. 6. 2012
<i>G. nepalensis</i>	DH-169	India	18. 7. 2014
<i>G. palmae</i>	DH-128	Spain	14. 7. 2013
<i>G. palmae</i>	DH-238	Spain	14. 7. 2013
<i>G. rhamni</i>	DH-01	Lebanon	2. 4. 2016
<i>G. rhamni</i>	DH-02	Lebanon	2. 4. 2016

<i>G. rhamni</i>	DH-08	Greece	22. 6. 2000
<i>G. rhamni</i>	DH-10	Greece	22. 6. 2000
<i>G. rhamni</i>	DH-11	Russia	4. 5. 2013
<i>G. rhamni</i>	DH-12	Russia	4. 5. 2013
<i>G. rhamni</i>	DH-20	Sweden	15. 7. 2006
<i>G. rhamni</i>	DH-21	Russia	27. 8. 2003
<i>G. rhamni</i>	DH-23	Sweden	15. 7. 2006
<i>G. rhamni</i>	DH-33	Austria	14. 8. 2016
<i>G. rhamni</i>	DH-34	Austria	14. 8. 2016
<i>G. rhamni</i>	DH-35	Austria	14. 8. 2016
<i>G. rhamni</i>	DH-49	Belarus	7. 7. 1990
<i>G. rhamni</i>	DH-51	Kazakhstan	26. 6. 2008
<i>G. rhamni</i>	DH-55	Kazakhstan	27. 7. 2014
<i>G. rhamni</i>	DH-59	Belarus	20. 8. 2000
<i>G. rhamni</i>	DH-63	Estonia	2. 7. 2016
<i>G. rhamni</i>	DH-64	Estonia	6. 7. 2016
<i>G. rhamni</i>	DH-65	Estonia	6. 7. 2016
<i>G. rhamni</i>	DH-67	Estonia	23. 7. 2016
<i>G. rhamni</i>	DH-68	Estonia	6. 7. 2016
<i>G. rhamni</i>	ZF-LY-004062	Czech Republic	16. 7. 2016
<i>G. rhamni</i>	ZF-LY-002534	Finland	-
<i>G. rhamni</i>	AP00017	Russia	2. 7. 2014
<i>G. rhamni</i>	ZF-LY-004181	Armenia	23. 6. 2006
<i>G. rhamni</i>	ZF-LY-004173	Morocco	26. 4. 2014
<i>G. rhamni</i>	DH-214	Lithuania	30. 6. 2007
<i>G. rhamni</i>	DH-292	Armenia	11. 7. 2010
<i>G. rhamni</i>	DH-295	Armenia	11. 7. 2010
<i>G. rhamni</i>	DH-296	Armenia	11. 7. 2010
<i>G. sp.</i>	DH-19	Iran	13. 5. 2016
<i>G. sp.</i>	DH-24	Greece	23. 6. 2008
<i>G. sp.</i>	DH-25	Greece	22. 6. 2008

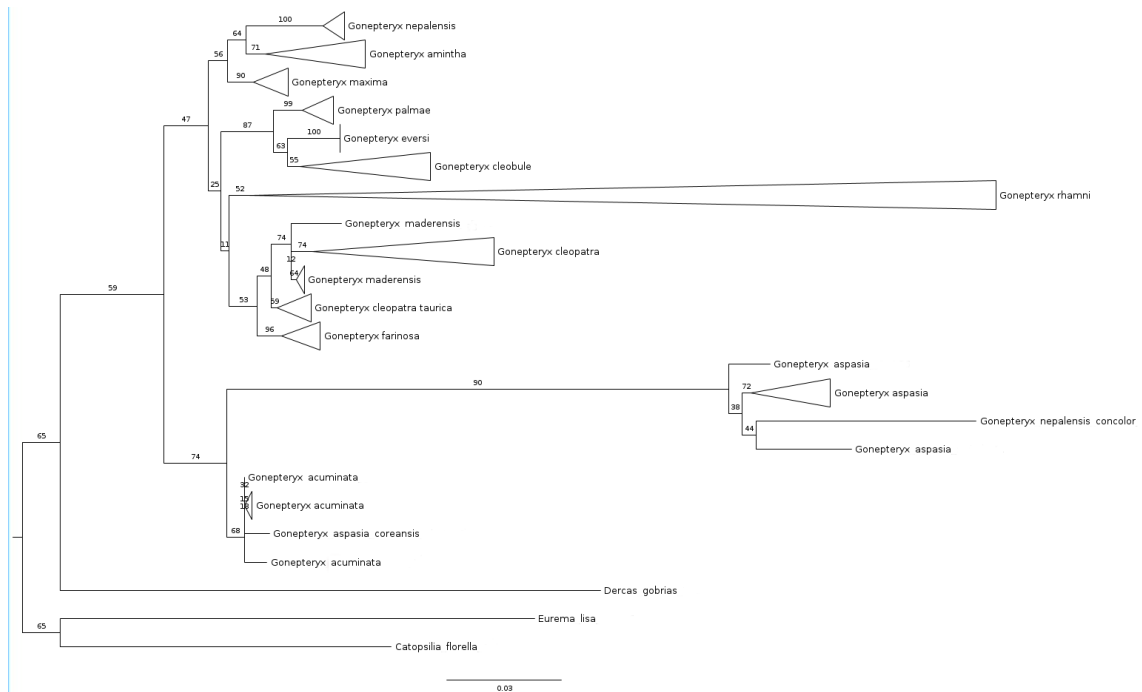
Appendix 2: Results of Maximum Likelihood: Phylogeny of the genus *Gonepteryx* based on COI, computed in RaxML.



Appendix 3: Results of Maximum Likelihood: Phylogeny of the genus *Gonepteryx* based on Wingless, computed in RaxML.



Appendix 4: Results of Maximum Likelihood: Phylogeny of the genus *Gonepteryx* based on both COI and Wingless, computed in RaxML.



Appendix 5: Phylogeny of the genus *Gonepteryx* based on both COI and Wingless, computed in BEAST 1.8.0.

